

## ASSESSMENT OF MICROPARTICULATE DRUG DELIVERY SYSTEM OF PROPRANOLOL HYDROCHLORIDE PREPARED BY MULTIPLE SOLVENT EMULSION TECHNIQUE

SATISH PANDAV, AMOLKUMAR LOKHANDE, JITENDRA NAIK\*

Department of Pharmaceutical Technology, University Institute of Chemical Technology, North Maharashtra University, Jalgaon - 425 001, India. Email: professorsnaik@gmail.com, satishpandav1985@rediffmail.com

Received: 06 July 2013, Revised and Accepted: 05 Aug 2013

### ABSTRACT

**Objective:** In the current work the sustained release microparticles of an antihypertensive drug propranolol hydrochloride was prepared due to its short half life and high water solubility.

**Method:** The sustained release microparticles of propranolol hydrochloride with ethyl cellulose of different ratios 1:1, 1:2, 1:3 and 1:4 (wt/wt) was prepared using different surfactants like polyvinyl alcohol and Tween 80 through the W/O/W double emulsion solvent evaporation method.

**Results:** The effect of polymer and surfactant concentration on the particle size, encapsulation efficiency and the drug release rate were studied. The obtained particles were characterized by Motic microscope, FE-SEM and FT-IR. *In vitro* release study was carried out using eight station dissolution test apparatus and UV- visible spectrophotometer at 289 nm. This study revealed that the maximum encapsulation efficiency was 78.22±0.59 %, no drug-polymer interaction and drug released from the microparticles was up to 12 h.

**Conclusion:** From obtained results it was concluded that this dosage form maintained the drug level in therapeutic window which may help to minimize the side effects, number of dosage and improve patient compliance.

**Keywords:** Propranolol hydrochloride, Ethylcellulose, Encapsulation efficiency, Sustained Release, Drug Release kinetics.

### INTRODUCTION

Microencapsulation is defined as the application of an emaciated coating of individual core materials that have a random particle size range from 1 to 1000  $\mu\text{m}$  [1]. Propranolol hydrochloride [1-(isopropylamino)-3-(1-naphthoxy)-2-propanol hydrochloride] is a nonselective beta-adrenergic receptor blocking agent with a biological half life of about 3-5 hours [2]. Propranolol hydrochloride is commercially available in the form of conventional formulations with dose 10 to 40 mg in 3-4 times daily. It is also indicated in the long term management of patients with angina pectoris. However, Propranolol hydrochloride usefulness is limited due to its short half-life. Hence, it requires three-four times a day dosing which produce the patient's noncompliance. To reduce the frequency of dose administration and to improve patient compliance, a controlled/sustained release formulation is popular [3]. Microparticles also offer advantages such as limiting fluctuate within a therapeutic range, reduction in side effects, decreased dose frequency and hence improved patient compliance [4,5]. Ethylcellulose, a non-biodegradable and biocompatible polymer is one of the extensively studied encapsulating materials for the controlled release of pharmaceuticals [6] was selected as the retardant material for Propranolol hydrochloride. Several researchers have investigated the utilization of ethylcellulose as a polymer to microencapsulate a drug by coacervation phase separation technique, emulsion solvent evaporation technique and spherical crystallization technique [7]. Generally, the solvent evaporation method is inappropriate for the microencapsulation of water soluble drugs as the associated efficiency of drug entrapment within the microparticles is low. This is primarily due to drug loss from the polymeric phase to the aqueous phase prior to solidification of the microparticles [8,9]. The purpose of the present work was to prepare and evaluate oral sustained release microparticulate drug delivery system of Propranolol hydrochloride using ethyl cellulose by W/O/W double emulsion solvent evaporation method with high entrapment capacity and extended release. Various parameters such as a drug-polymer ratio, surfactant concentration were optimized to maximize the entrapment. These microparticles were evaluated for encapsulation efficiency, drug content and *in vitro* drug release. Drug-polymer interactions in the solid state were studied by Fourier transform infrared spectroscopy (FT-IR), the size distribution and surface morphology were evaluated by Motic microscope and FE-SEM respectively.

### MATERIALS AND METHOD

#### Materials

Propranolol hydrochloride was gifted from Micro Advanced Research Center, Pvt. Ltd., Bangalore and Ethyl Cellulose was received as a gift sample from Colorcon Asia Pvt. Ltd., Goa. Gelatin, extra pure was purchased from Himedia Laboratories Pvt. Ltd. Mumbai. PVA (MW 1, 25,000), Tween 80, Dichloromethane and n-hexane were procured from Merck Specialities Pvt. Ltd., Mumbai. Dissolution medium was prepared by using triple distilled water filtered with 0.22  $\mu\text{m}$  membrane filter.

#### Method

##### Preparation of microparticles

Weighed amount of ethyl cellulose was dissolved in dichloromethane. Deionized aqueous drug solution was added in above polymeric organic solution to preparing a primary emulsion (W/O) with constant stirring at 500 rpm for 15 min. Gelatin (% w/v) was added in aqueous solution of drug to improve the viscosity of the solution. This primary emulsion was added drop by drop to the aqueous solution of pH 12 containing polyvinyl alcohol (% w/v) or Tween 80 (% w/v) as a surfactant with 1000 rpm constant stirring for 2 h using a lab stirrer (Remi Elektrotechnik Ltd. Mumbai). The n-hexane was added to harden the formed microparticles and the stirring was continued for further 1 h. The resulting microparticles were collected by filtration. The product was then dried at room temperature for 24 h and stored in desiccators for 74 h. All batches were prepared in triplicate [10, 11].

##### Particle size measurement

Motic microscope (B1 series, Motic, China) was used to determine the size microparticles. A drop of suspended microparticles was spread on a glass slide and observed at 40x and 100x resolution power using a microscope installed with a digital camera and image analysis software. Three areas on the slide containing about 100 particles were randomly snapped and used to determined the average particle size range [12].

##### Determination of encapsulation efficiency

Weighed quantities of respective samples were added in dichloromethane to dissolve polymer and then add the required

volume of double distilled water to extract the drug in it and precipitated the polymer. Finally this drug containing dichloromethane-aqueous solution was stirred until complete evaporation of dichloromethane. The remained aqueous solution of the drug was filtered through 0.45 µm pore size Whatman filter paper and analyzed spectrophotometrically at wavelength 289 nm using UV- visible spectrophotometer (UV- 1800 Shimadzu Co. Ltd., Japan). The drug encapsulation efficiency was calculated using following equation 1:

$$EE (\%) = D_m \times 100 / D_t \dots\dots\dots (1)$$

Where  $D_t$  is the theoretical amount of drug loaded to polymer solution and  $D_m$  is the practical amount of drug encapsulated in the prepared microparticles [13]. Determinations of encapsulation efficiency were run in triplicate.

#### Surface morphology

The shape and surface properties of microparticles were investigated by using Field Emission-Scanning Electron Microscopy (S4800, Hitachi, Japan). An appropriate sample was mounted on stub, using double sided adhesive carbon tapes and observed for morphology at acceleration voltage.

#### Fourier transform infrared spectroscopy

Propranolol hydrochloride, Ethyl cellulose and drug loaded microparticles were mixed with KBr and pressed to disk. Infrared spectra of the samples were scanned in the range from 400 to 4000  $\text{cm}^{-1}$  and recorded on a Fourier transform infrared spectrometer (Shimadzu Co. Ltd., Singapore) to study the possible interaction between propranolol hydrochloride and ethyl cellulose. All measurements were taken at room temperature. The spectra of water and KBr were subtracted from the sample spectrum and the procedure was done under nitrogen gas to prevent humidity interference [14].

#### In-vitro drug release study

*In vitro* dissolution studies were performed using USP Type II dissolution test apparatus (Paddle) at 75 rpm (Electrolab, India). The dissolution medium was used as double distilled deionized water. Accurately weighed samples of the microparticles were added to dissolution medium and withdraw 5mL dissolution

medium by 0.45µm attached Whatman filter paper syringe at preselected time intervals up to 12 hours monitored progress of the dissolution. Same volume of dissolution medium was replenished after each sampling. The absorbance of the withdrawn samples was measured at 289 nm against blanks. The concentration of Propranolol hydrochloride in test samples was corrected and calculated using a regression equation of the calibration curve. The dissolution studies were carried out in triplicate and the mean values were plotted as percentage cumulative drug release versus time.

#### In-vitro release kinetics

The *in-vitro* release profiles were fitted to zero order, first order, Higuchi square root and Hixon-Crowell model. Regression coefficients ( $R^2$ ) were determined from the slope of the all obtained plots. Mechanism of drug release according to Korsmeyer-Peppas model was evaluated by fitted first 60% of drug release in its equation and release exponent "n" was calculated from plot Log cumulative % drug release vs Log time [15].

#### RESULTS AND DISCUSSION

In this work the effect of drug-polymer ratio and surfactant concentrations on particle size, entrapment efficiency and release pattern of propranolol hydrochloride from ethyl cellulose microparticles prepared by solvent evaporation method were examined. Propranolol was highly soluble in water therefore to reduce its solubility in external phase the pH of the external aqueous solution was increased up to 12. Due to decreased in solubility of Propranolol HCl in external water phase the drug loss was reduced and enhance the encapsulation efficiency [16]. Gelatin was used as an emulsifier and viscosity enhancer of the primary emulsion. Ethyl cellulose, also acts as a self emulsifier to stabilize this emulsion. Dichloromethane is less soluble and having more interfacial tension with water, therefore formed stable multiple emulsion (W/O/W). The emulsion droplets containing propranolol hydrochloride get transformed into solid state due to ethyl cellulose hydrophobic property and dichloromethane evaporation during stirring. This evaporation rate was maintained by stirring at 1000 rpm and 25 °C temperature. Table 1 shows the influence of drug-polymer ratio, surfactant and its concentration on particle size, percentage yield and encapsulation efficiency of different formulation trials.

Table 1: Effect of content concentration variables on Propranolol HCl loaded microparticles.

Formulation Code	Drug:Polymer Ratio	Surfactant concentration	Yield (%)	Encapsulation Efficiency	Particle Size (µm)
F1	1:1	0.2% PVA	72.46±0.42	48.56±0.41	100-150
F2	1:2	0.2% PVA	75.46±0.43	63.75±0.31	150-200
F3	1:3	0.2% PVA	78.44±0.43	66.79±0.49	170-240
F4	1:4	0.2% PVA	78.41±0.57	73.74±0.44	230-300
F5	1:1	0.5% PVA	69.57±0.60	52.61±0.43	50-100
F6	1:2	0.5% PVA	77.58±0.52	66.72±0.51	100-150
F7	1:3	0.5% PVA	82.56±0.53	72.04±0.73	125-200
F8	1:4	0.5% PVA	82.83±0.65	78.22±0.59	200-250
F9	1:4	0.2% T80	68.60±0.73	66.22±0.79	280-400
F10	1:4	0.5%T80	79.48±0.51	69.99±0.64	250-300

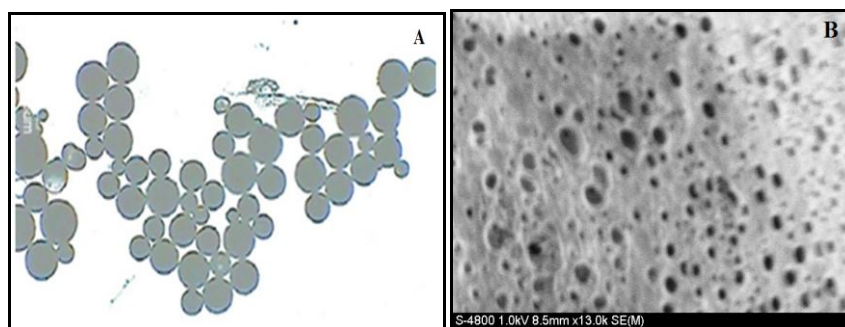


Fig. 1: (a) Motic microscope image and (b) Surface morphology of recovered microparticles.

The obtained results concluded that the particle sizes were directly proportional to ethyl cellulose concentration may be due to increased in viscosity of the internal phase [15], and inversely with the surfactant concentration may be due to the formation of new surfaces for the small emulsion globules [17]. The Motic microscope image is shown in Fig. 1a. Encapsulation efficiency was directly proportional to both the polymer and surfactant concentration. Because the polymer concentration may help to enhance the holding capacity for drug and surfactant decreased the interfacial tension between aqueous drug solution and polymeric organic solution [18].

The particles formulated by Tween 80 were less encapsulated and more size than the formulations prepared with polyvinyl alcohol because PVA has a better propensity to drift towards the surface of microparticles and stabilized it more efficiently. All recovered microparticles were spherical in shape and slightly porous in nature (Fig. 1b). The porous structure was responsible for the sustained release of drugs.

Drug polymer interaction was determined by comparing the IR spectra of propranolol hydrochloride loaded ethyl cellulose microparticles with the IR spectrum of pure drug.

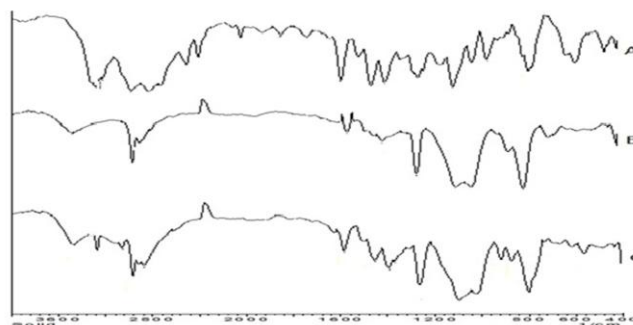


Fig. 2: FT-IR Spectra of pure Propranolol hydrochloride (A), unloaded (B), and optimized formulation (C).

As shown in Figure 2, propranolol hydrochloride (A) gives peaks in IR spectrum nearby at 2965  $\text{cm}^{-1}$  due to the presence of a secondary amine group, 3283  $\text{cm}^{-1}$  due to the hydroxyl group (secondary), the aryl alkyl ether shows a stretching band at 1267.27  $\text{cm}^{-1}$  and the peak at 798  $\text{cm}^{-1}$  due to a-substituted naphthalene [19]. Frequencies of functional groups of pure drug remained intact or overlapped by polymer in physical mixture containing polymer.

Hence, there was no major interaction between the drug and polymer used in the study.

Figure 3. Illustrate *in vitro* release of selected formulations in double distilled water, by representing the cumulative percentage of propranolol hydrochloride released with respect to the amount of drug encapsulated. The formulations were selected on the basis of higher encapsulation.

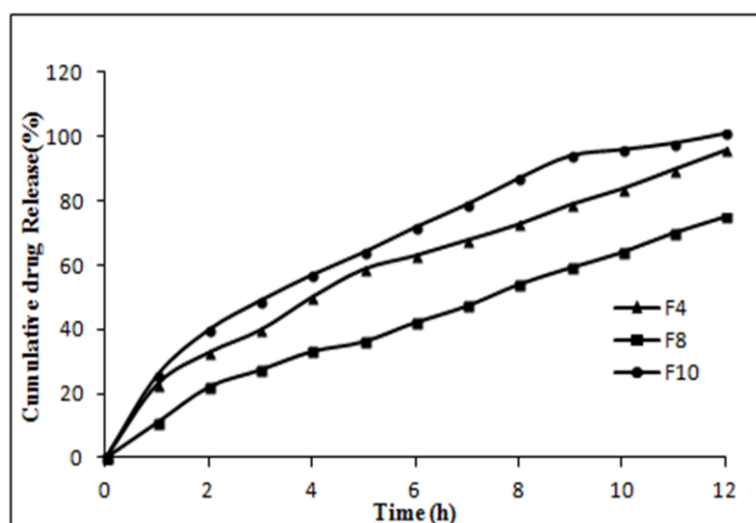


Fig. 3: Release profile of propranolol hydrochloride from some selected formulations

The formulation F8 was more sustained than formulation F4 because the maximum number of particles may extend the time to release the drug from F8 formulation. In F8 formulation the 0.5%, w/v PVA surfactant was used which gives small size particles than F4 formulation (0.2%, w/v PVA). So, the same weight of particles of both formulations (F4 and F8) contained maximum number of particles in F8 formulations. When we compared the effect of surfactant on dissolution profile it was concluded that the formulations prepared by using PVA was more sustained than

Tween 80 used formulations. Because PVA may stabilize the surface more effectively and encapsulates the drug at the core of particles. At the end of 12 hours F8 formulation released  $74.88 \pm 0.39$ , F4 released  $95.92 \pm 0.65$  and F10 released  $101.04 \pm 0.52$  per cent Propranolol hydrochloride. Formulation F10 and F4 released  $25.79 \pm 0.87$  %,  $23.02 \pm 0.56$  % drug at the first hour, may be due to the drug present at the surface of particles. The burst release effect may be due to the adsorption of the drug on the surface of the particles or due to concentrating the drug at the surface of the

particles because insufficient concentration or ineffectual surfactant unable to encapsulate drug at the core of particles and drug moved towards the interface of both phases. In F10 formulation the reason was Tween 80 not efficiently stabilized the surface of the particles during solidification step and in F4 formulation the PVA was in deficient concentration to stabilize the surface of the particles. Therefore the drug was not encapsulated in the core of the particle but stabilized or adsorbed at the surface of the particles.

The release kinetics of all selected formulations explained in Table 2. The obtained results revealed that the most sustained F8 formulation best fitted in zero order kinetics. The zero order rates describe the system where the drug release rate is independent of time and its concentration within microparticles. In this kinetics drug releases constantly over time, minimizing side effects and maintained drug concentration within the therapeutic window. Therefore preserve the drug level in blood throughout the delivery period [20].

**Table 2: *In vitro* release kinetics of selected formulations**

Formulations	Zero Order	First Order	Higuchi	Hixon-Crowell	n-value
F4	0.9847	0.8947	0.9955	0.9664	1.7192
F8	0.9944	0.9740	0.9833	0.9880	1.3735
F10	0.9630	0.9290	0.9923	0.8613	1.7419

Formulations F4 and F10 follows Higuchi model. Many controlled release products are designed on the principle of implanting the drug in a porous matrix. The release of solid drug from a granular matrix involves the simultaneous penetration of surrounding liquid, dissolution of the drug, and leaching out of the drug through interstitial channels or pores. The volume and length of opening were also accounted in the diffusion theory [21]. The mechanism of drug release according to Korsmeyer-Peppas model results (n-value) revealed that the overall solute diffusion mechanism follows Super case-II transport. Case-II relaxational release is the drug transport mechanism associated with stresses, state transition in polymers which swell in water, polymer Disentanglement and erosion [22-24]. From all above evaluations it was concluded that the prepared microparticles were successfully sustained for 12 h.

#### CONCLUSION

From the above results it was concluded that propranolol hydrochloride was successfully encapsulated into ethylcellulose microparticles using W/O/W double emulsion solvent evaporation method. Polyvinyl alcohol was more suitable surfactant than Tween 80 with a concentration of 0.5%, w/v. The 1:4 drug-polymer ratio obtained highest encapsulation and sustained the Propranolol hydrochloride for 12 h. It follows zero order release kinetics therefore this dosage form maintain the drug level in therapeutic window which may help to minimize the side effects, number of dosage and improve patient compliance.

#### ACKNOWLEDGMENT

Authors are very much thankful to Defence Research and Development Organization, New Delhi for granting financial support for this valuable research work and Micro Advanced Research Center, Pvt. Ltd., Bangalore for providing a gift sample of Propranolol hydrochloride as well as Colorcon Asia Pvt. Ltd., Goa for Ethyl cellulose.

#### REFERENCES

- Nordstierna L, Abdalla AA, Nordin M, Nyden M. Comparison of release behavior from Microcapsules and microspheres. *Prog Org Coat* 2010; 69:49-51.
- Derle D, Joshi O, Pawar A, Patel J, Jagadale A. Formulation and Evaluation of Buccoadhesive Bi-layer Tablet of Propranolol Hydrochloride. *Int J Pharm Pharm sci* 2009; 1(1): 206-212.
- Sexena L, Singh RK, Diwivedi A, Khan I, Singh A, Raghvendra. Preparation and Evaluation of Microspheres of Propranolol Hydrochloride Using Eudragit RL as the Material. *American-Eurasian J Sci Res* 2011; 6 (2):58-63.
- Davis. The design and evaluation of controlled release systems for the gastrointestinal tract. *J Control Release* 1985; 2: 69-77.
- Ritschel WA. Biopharmaceutics and Pharmacokinetic aspects in the design of controlled release peroral drug delivery systems. *Drug Deve Ind Pharm* 1989; 15: 1073-1103.
- Murtaza G. Ethylcellulose microparticles: a review. *Acta Pol Pharm* 2012; 69 (1): 11-22.
- Das MK, Rama Rao K. Evaluation of zidovudine encapsulated ethylcellulose microspheres prepared by water-in-oil-in-oil (w/o/o) double emulsion solvent diffusion technique. *Acta Pol Pharm-Drug Research* 2006; 63 (2): 141-148.
- Davis S, Kirsten J. Contribution of process variables to the entrapment efficiency of Propranolol hydrochloride within ethylcellulose microspheres prepared by Solvent Evaporation Method as evaluated using a Factorial design. *Int J Pharm* 1996; 131: 25-31.
- Bodmeier R, Mc Ginity JW. Solvent selection in the preparation of poly (d, l) lactide microspheres prepared by the solvent evaporation method. *J Pharm Pharmacol* 1988; 43: 179-186.
- Basu SK, Adhiyaman R. Preparation and Characterization of Nitrendipine loaded Eudragit RL 100 Microspheres Prepared by an Emulsion-Solvent Evaporation Method. *Tropical J Pharmaceutical Res* 2008; 7 (3): 1033-1041.
- Naik JB, Lokhande AB, Mishra S, Kulkarni RD. Development of sustained release micro/nanoparticles using different solvent emulsification technique: A review. *Int J Pharm Bio Sci* 2012; 3(4) : 573-590.
- Lokhande AB, Deshmukh TA, Patil VR. Fabrication and *in vitro* evaluation of metformin entrapped sustained released ammonio methacrylate copolymer microparticles prepared by non aqueous emulsion solvent evaporation technique. *Int J Pharm Bio Sci* 2013; 4(3): 444-453.
- Jaganathan KS and Singh P. Development of a single dose tetanus toxoid formulation based on polymeric microspheres: a comparative study of poly (d, l-lactic-co-glycolic acid) versus Chitosan microspheres. *Int J Pharm* 2005; 294: 23-32.
- Hamishehkar H, Emami J, Najafabadi AR, Gilani K, Minaiyan H, Mahdavi M, Nokhodchi A. The effect of formulation variables on the characteristics of insulin-loaded poly (lactic-co-glycolic acid) microspheres prepared by single phase oil in oil solvent evaporation method. *Colloids Surf B Biointerfaces* 2009; 74: 340-349.
- Lokhande AB, Mishra S, Kulkarni RD, Naik JB. Preparation and characterization of repaglinide loaded ethyl cellulose nanoparticles by solvent diffusion technique using high pressure homogenizer. *Journal of Pharmacy Research* 2013;7:421-426.
- Perez MH, Zinutti C, Lamprecht A, Ubrich N, Astier A, Hoffman M, Bodmeier R, Maincent P. The preparation and evaluation of poly (e-caprolactone) microparticles containing both a lipophilic and a hydrophilic drug. *J controlled Release* 2000; 65:429-438.
- Lee SC, Oh JT, Jang MH, Chung S. Quantitative analysis of polyvinyl alcohol on the surface of poly (D, L-lactide-co-glycolide) microparticles prepared by solvent evaporation method: effect of particle size and PVA concentration. *J controlled Release* 1999; 59:123-132.
- Jayanthi B, Manavalan R, Manna PK. Effect of process parameters on the physiochemical properties of aceclofenac loaded microparticles. *Int J Pharm Pharm sci* 2012; 4(4): 471-478.
- Dabbagh MA, Ford JL, Rubinstein MH, Hogan JE, Rajabi-Siahboomi AR. Release of Propranolol hydrochloride from matrix tablets containing sodium carboxymethylcellulose. *Pharm Dev Tech* 1999; 4:313-324.

20. Lokhande AB, Mishra S, Kulkarni RD, Naik JB. Influence of different viscosity grade ethylcellulose polymers on encapsulation and In vitro release study of drug loaded nanoparticles. *Journal of Pharmacy Research* 2013; 7:414-420.
21. Higuchi WI. Diffusional models useful in biopharmaceutics drug release rate processes. *J Pharm Sci* 1967; 56: 315-324.
22. Korsmeyer RW, Lusting SR, Peppas NA. Solute and penetrant diffusion in swellable polymers. I. Mathematical modeling. *J polym Sci Polym Phys Ed* 1986a; 24: 395-408.
23. Korsmeyer RW, Von ME, Peppas NA. Solute and penetrant diffusion in swellable polymers. II. Verification of theoretical models. *J polym Sci Polym Phys Ed* 1986b; 24: 409-434.
24. Cox PJ, Khan KA, Munday DL, Sujja AJ. Development and evaluation of a multiple-unit oral sustained release dosage form for S (+) -ibuprofen: preparation and release kinetics. *Int J Pharm* 1999; 193: 73-84.