

ISSN- 0975-7066

Vol 8, Issue 3, 2016

**Original Article** 

# ANTIOXIDANT ACTIVITY OF OYSTER MUSHROOM (*PLEUROTUS FLORIDA* [MONT.] SINGER) AND MILKY MUSHROOM (*CALOCYBE INDICA* P AND C)

## MADHAIYAN PRABU\*, RENGANATHAN KUMUTHAKALAVALLI

Department of Biology, Gandhigram Rural Institute-Deemed University, Gandhigram 624302 Dindigul (TN), India Email: sma.subi@gmail.com

#### Received: 12 Mar 2016, Revised and Accepted: 10 Jun 2016

## ABSTRACT

Objective: To evaluate the antioxidant activity of tropical edible mushrooms namely Pleurotus florida and Calocybe indica.

**Methods:** Antioxidant potential was evaluated by using various antioxidant assays such as DPPH free radical scavenging, hydroxyl radical scavenging, nitric oxide radical scavenging, and superoxide radical scavenging activities as well as lipid peroxidation inhibiting assay, reducing power assay, ferric reducing antioxidant power (FRAP), metal chelating activity, phospho-molybdenum reduction assay and anti-haemolytic activity.

Results: The results obtained from this antioxidant study strongly suggest that Pleurotus florida and Calocybe indica have significant antioxidant activity.

**Conclusion:** Edible mushrooms *Pleurotus florida* and *Calocybe indica* are having significant antioxidant activity, could serve as easily accessible natural food rich in antioxidant which may enhance the immune system against oxidative damage and may be utilized as the potential sources of therapeutic agents.

Keywords: Antioxidant, Pleurotus florida, Calocybe indica, Tropical edible mushrooms, Therapeutic agents

© 2016 The Authors. Published by Innovare Academic Sciences Pvt Ltd. This is an open access article under the CC BY license (http://creativecommons.org/licenses/by/4.0/)

#### INTRODUCTION

Mushrooms are eukarvotic, non-photosynthetic organisms that form characteristic fruiting bodies. They have been used by human beings for thousands of years as food and medicine [1]. The increasing consumption of mushrooms can be attributed not only to the pleasant flavour and aroma of edible mushrooms but also to their vitamins and protein contents as well as the molecule that include antioxidants and natural products which are free from pesticides [2]. Mushrooms are now considered as potential therapeutics and preventive agents that may ensure the wellness of humans. As a result, mushroom cultivation has been increased tremendously throughout the world during the last few decades [3]. Mushrooms have been reported to have significant pharmacological effects such as regulation of biorhythm, maintenance of homeostasis, prevention and cure for various diseases such as cancer, cerebral stroke, heart diseases and improvement of life. They have also been demonstrated to contain effective biomolecules with hypolipidemic, antithrombotic, hypotensive, anti-inflammatory and other applications [4]. The present study aims to evaluate the antioxidant activities of the extract of edible mushrooms Pleurotus florida and Calocybe indica.

#### MATERIALS AND METHODS

The fruiting bodies of *Pleurotus florida* and *Calocybe indica* were obtained from Mushroom Unit, Department of Biology, The Gandhigram Rural Institute–Deemed University, Gandhigram (TN), India. Sample preparation [5] and antioxidant activities such as DPPH free radical scavenging [6], hydroxyl radical scavenging [7], nitric oxide radical scavenging [8], superoxide radical scavenging [9], lipid peroxidation inhibiting assay [10], reducing power assay [11], ferric reducing antioxidant power (FRAP) assay [12], metal chelating activity [13], phospho-molybdenum reduction assay [14] and anti-haemolytic activity [15] of methanol extract were carried out.

### Statistical analysis

The results were expressed as mean values and standard deviation (SD). Linear regression analysis was used to calculate IC50 value. Data were analyzed using One-Way Analysis of Variance (ANOVA)

followed by Turkey's multiple comparison post hoc tests using SPSS software 16.0 versions. Values of p<0.05 were considered as statistically significant.

#### RESULTS

The antioxidant capacity of *Pleurotus florida* and *Calocybe indica* was determined by the DPPH method and the results were presented in fig. 1. Different concentrations of P. florida and C. indica (200-1000 µg/ml) showed maximum DPPH radical scavenging activity of 37.04±0.15 and 28.04±0.41 % at 1000 µg/ml respectively. Results showed the percentage of inhibition in a dose-dependent manner. The IC<sub>50</sub> value of *P. florida* and *C. indica* were found to be 413.28±5.87 µg/ml and 588.40±11.85 µg/ml respectively. The results of hydroxyl radical scavenging activity were recorded in fig. 2. The hydroxyl radical scavenging effects of extracts of P. florida and C. indica using deoxyribose assay in the different concentrations (200-1000 µg/ml) were investigated. The strongest hydroxyl radical scavenging activity was observed in C. indica (65.41±0.65 % at 1000 µg/ml) than *P. florida* (46.99±2.58 % at 1000 µg/ml). The IC<sub>50</sub> value of P. florida and C. indica was found to be 220.70±6.0 µg/ml and 148.23±1.01 µg/ml respectively.

Fig. 3 depicts the nitric oxide radical scavenging activity of the extracts. Different concentrations of Pleurotus florida and Calocybe indica (200-1000 µg/ml) showed 21.90±0.88 % and 23.13±1.32 % inhibition at the concentration of 1000 µg/ml. Concentrations required for 50% inhibition (IC<sub>50</sub>) of nitric oxide radical scavenging activity in P. florida and C. indica were 893.44±28.14 and 781.63±20.83 µg/ml respectively. P. florida and C. indica were found to scavenge superoxide generated by photoreduction of riboflavin (fig. 4). The IC<sub>50</sub> value of *P. florida* and *C. indica* were found to be  $357.29\pm8.93$  µg/ml and  $441.92\pm7.81$  µg/ml respectively. The inhibitory effects of P. florida and C. indica on lipid peroxidation inhibition were increased with increasing concentration. The inhibitory effect of the different concentrations of extract such as 200-1000 µg/ml of *P. florida* and *C. indica* were determined using the liver homogenate model and the results were recorded in fig. 5. The results of lipid peroxidation inhibition were found to be 80.0±4.01 % at 1000 µg/ml in P. florida and 87.59±0.32 % at 1000

 $\mu$ g/ml in *C. indica*. The IC<sub>50</sub> value of *P. florida* and *C. indica* was found to be 17.44±0.64  $\mu$ g/ml and 16.06±0.25  $\mu$ g/ml respectively.

Fig. 6 summarizes that the reducing power of the extracts of *Pleurotus florida* and *Calocybe indica* was found to be excellent which steadily increased in direct proportion to the increasing concentrations of the extract. The reducing power inhibition percentages were found to be  $0.43\pm0.007$  % in *P. florida* and  $0.43\pm0.005$  % in *C. indica*.

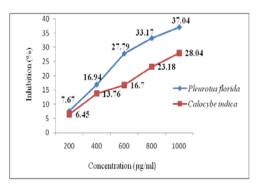


Fig. 1: DPPH radical scavenging activity

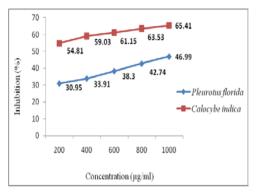


Fig. 2: Hydroxyl radical scavenging activity

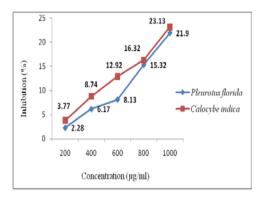


Fig. 3: Nitric oxide radical scavenging activity

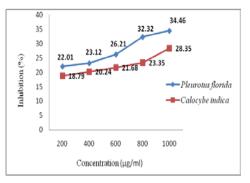


Fig. 4: Superoxide radical scavenging activity

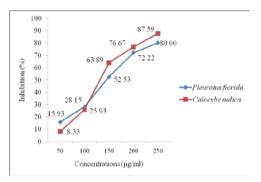


Fig. 5: Lipid peroxidation assay

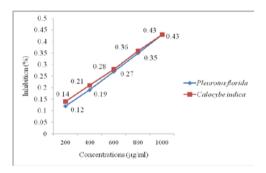


Fig. 6: Reducing power assay \*Values are mean±SD (*n* = 3) (*p*<0.05)

In order to examine the reducing power, the reduction of Fe<sup>3+</sup>to Fe<sup>2+</sup>was investigated in the *Pleurotus florida* and *Calocybe indica* mushroom extract and the result is shown in table 1. The absorbance of the reaction mixtures at 593 nm was found as 59.65±0.46 m. mol [Fe (II]]/g and 57.38±0.37 m. mol [Fe (II]]/g in *P. florida* and *C. indica* respectively. Metal chelating activity was found to be higher in *C. indica* (0.97±0.05 mg EDTA E/g extract) than *P. florida* (0.77±0.08 mg EDTA E/g extract) (table 1). The antioxidant capacity (phospho-molybdenum reduction assay) of the extract of *P. florida* (197.26±1.19 %) was recorded to be higher than *C. indica* (190.64±0.40 %) (table 1).

Table 1: Ferric reducing antioxidant power	· (FRAP) assay, Metal chelating	ng activity and phospho-molybdenum as	sav

Sample	FRAP mmol [Fe (II)]/g extract	Metal chelating activity (mg EDTA Eq/g extract)	Phosphomolybdenum (mg ascorbic acid Eq/g extract)
Pleurotus florida	59.65±0.46 %	0.77±0.08 %	197.26±1.19 %
Calocybe indica	57.38±0.37 %	0.97±0.05 %	190.64±0.40 %

Values are mean $\pm$ SD (*n* = 3) (*p*<0.05). In the present study, extract of *Pleurotus florida* (20.23 $\pm$ 3.33 %) and *Calocybe indica* (10.21 $\pm$ 3.66 %) exhibited potent anti-haemolytic activity (table 2).

Table 2: Anti-haemolytic activity

Sample	Concentration(µg/ml)	Percentage activity (%)	
Pleurotus florida	250	10.21±3.66 %	
Calocybe indica	250	20.23±3.33 %	

Values are mean $\pm$ SD (n = 3) (p < 0.05).

#### DISCUSSION

The Oyster and milky mushrooms produce a very impressive array of antioxidant compounds and have the potential to lower the risk of diseases [16, 17]. The free radical scavenging ability of these extracts of *Pleurotus florida* and *Calocybe indica* were found to be on the high side. This result is well supported by Blois [6] and Gezer *et al.* [18]. Their findings revealed that cysteine, glutathione, ascorbic acid,  $\alpha$ -tocopherol, polyhydroxy aromatic compounds and aromatic amines reduce and decolorize  $\alpha$ ,  $\alpha$ -diphenyl- $\beta$ -picrylhydrazyl by their hydrogen donating ability. The antioxidative activities dose dependency and associated it the presence of reductones are reported to be the terminators of free radical chain reactions [19].

The hydroxyl radical is an enormously reactive free radical created in biological systems and has been concerned with an extremely harmful species in free radical pathology able to damage nearly each molecule found in living cells. This radical has the power to bond nucleotides in DNA and cause strand rupture, which leads to cause cytotoxicity carcinogenesis and mutagenesis. The ability of extracts to quench hydroxyl radicals seems to be the good scavenger of active oxygen species, thus reducing the rate of the chain reaction [20-24].

Nitric oxide is an important chemical mediator generated by endothelial cells, macrophages, neurons, etc., and is involved in the regulation of various physiological processes. Excess concentration of nitric oxide is associated with several diseases. Oxygen reacts with the excess nitric oxide to generate nitrite and peroxynitrite anions which act as free radicals [25]. Similar antioxidant activity for nitric oxide in *Pleurotus florida* and *Calocybe indica* were reported by different research groups [26-31].

Superoxide anion is a reduced form of molecular oxygen created by receiving one electron. Superoxide anion is an initial free radical formed from mitochondrial electron transport systems. Mitochondria generate energy using 4-electron chain reactions reducing oxygen to water. Some of the electrons escaping from the chain reaction of mitochondria directly react with oxygen and form superoxide anion. It plays an important role in the formation of other reactive oxygen species such as hydrogen peroxide, hydroxyl radical or singlet oxygen in living systems [24, 29, 32, 33].

The lipid peroxidation has been broadly defined as the oxidative deterioration of polysaturated lipids. Peroxyl and hydroxyl radicals are important agents that mediate lipid peroxidation, thereby damaging cell membranes. A number of toxic compounds are generated during this process of lipid peroxidation. Thiobarbituric acid reactive substances (TBARS) are produced as by-products of lipid peroxide that occurs in the hydrophobic core of biomembranes. A substance may act as an antioxidant due to its ability to reduce ROS by donating hydrogen atom [22, 24, 34-36]. The effect of mushroom extracts on lipid peroxide showed significant inhibition of TBARS formation. The present finding strongly suggests that the use of the mushroom extracts prevent lipid peroxide and this arrests membrane damage.

The antioxidant activities of certain mushroom extracts have been related to their reducing potential. The reducing potential of the extract of *Pleurotus florida* and *Calocybe indica* was evaluated using ferric reducing assay. The reducing potency is generally associated with the presence of substances called reductones, which exert antioxidant action by breaking the free radical chains *via* hydrogen atom donation. Reductones are reported to prevent peroxide formation by reacting with certain precursors of peroxides. In this assay, the presence of reductants in the samples would result in the reducing of Fe<sup>3+</sup>to Fe<sup>2+</sup>by donating the electron. The amount of Fe<sup>2+</sup>complex can be measured by measuring the formation of Perl's Prussian blue at 700 nm. Increasing absorbance indicates an increase in reductive ability [37-43].

Further, the reducing capacity was investigated by measuring  $Fe^{3+}to$   $Fe^{2+}conversion$  and serve as a significant indicator of its potential antioxidant activity. The antioxidant activities of putative antioxidants have been attributed to various mechanisms such as prevention of chain reaction, binding of transition metal ion catalysts, decomposition of peroxides, prevention of continued proton obstruction and radical scavenging [25].

Antioxidants inhibit the interaction between metal and lipid through the formation of insoluble metal complexes with a ferrous ion. The iron chelating capacity test measures the ability of antioxidants to compete with ferrozine in chelating ferrous ion [17]. Transition metals have been proposed as the catalysts for the initial formation of radicals. Chelating agents may stabilize transition metals in living systems and inhibit generation of radicals, consequently reducing free radical induced damage. To estimate the antioxidant potential of *Pleurotus florida* and *Calocybe indica* mushroom extracts, their chelating activity was evaluated against  $Fe^{2*}$ . Ferrozine quantitatively forms complexes with  $Fe^{2*}$ . The chelating effects of the *P. florida* and *C. indica* mushroom extracts were found to be excellent.

Total antioxidant capacity by phospho-molybdenum method assay is based on the reduction of Mo (VI) to Mo (V) by the sample analyze and the subsequent formation of green phosphate/Mo (V) complex at acidic pH. The phospho-molybdenum method is quantitative since the total antioxidant activity is expressed as the number of equivalents of ascorbic acid [14]. Phosphomolybdenum assay with the methanolic extracts of *Pleurotus florida* and *Calocybe indica* was determined. Comparatively, *P. florida* shows a better reduction of Mo than *C. indica*. The phospho-molybdenum assay results indicate that the methanolic extracts of *Grewia hirsuta* Vahl (Kalunnu) are more powerful antioxidant in the reduction of phospho-molybdenum complex [44].

Erythrocytes are considered as the major target for the free radicals owing to the presence of both high membrane concentration of polyunsaturated fatty acids (PUFA) and the oxygen transport associated with redox active haemoglobin molecules, which are potent promoters of activated oxygen species. The extent of haemolysis was found to be much greater when red blood cells were treated with hydrogen peroxide (toxicant). This could be attributed to the oxidizing nature of hydrogen peroxide with respect to the destruction of a cell membrane and subsequent liberation of haemoglobin from the cells. Mobilization of Fe2+by Ca2+via Fenton reaction is also caused due to hydrogen peroxide which further leads to the production of OH radicals. All these factors, in unison, cause deterioration of cell membrane, which may, perhaps, be the key episode of the lyses of the cell. Nevertheless, the antihaemolytic activity is the expression of collaborative action of the various antioxidant mechanisms which function in nature [45-47].

### CONCLUSION

The results obtained from this antioxidant study strongly suggest that the extract of *Pleurotus florida* and *Calocybe indica* has significant antioxidant activity, could serve as an easily accessible item of natural rich antioxidant food which may enhance the immune system against oxidative damage or it may be utilized as a potential source of therapeutic agent.

## **CONFLICT OF INTERESTS**

We declare that we have no conflict of interests

## REFERENCES

1. Wasser SP. Medicinal mushrooms as a source of antitumour and immunomodulating polysaccharides. Appl Microbiol Biotechnol 2002;60:258–74.

- Royse DJ. Forward to the Fifth International Conference on mushroom biology and mushroom products. Acta Edulis Fungi 2005;12:1–2.
- 3. Prabu M, Kumuthakalavalli R. Nutritional and phytochemical studies on *Pleurotus florida* (Mont.) Singer and *Calocybe indica* P and C. World J Pharm Res 2014;3:4907–13.
- Wong SP, Naidu M, David P, Bakar R, Vikineswary S. Neurogenerative potential of Lion's mane mushroom, *Hericium erinaceus* (Bull.: Fr.) Pers., (Higher Basidiomycetes), in the treatment of peripheral nerve injury. Int J Med Mushrooms 2012;14:427–46.
- Suffness M, Douros J. Drugs of plant origin. In: DeVita VT, Busch H. editors. Methods in Cancer Research, Academic Press: New York; 1979;26:73–126.
- 6. Blois MS. Antioxidant determinations by the use of a stable free radical. Nature 1958;26:1199–200.
- Klein BP, Perry AK. Ascorbic acid and Vitamin A activity in selected vegetables from different geographical areas of the United States. J Food Sci 1982;47:941–5.
- 8. Sreejayan N, Rao MNA. Nitric oxide scavenging activity *Curcuminoids*. J Pharm Pharmacol 1997;47:105–7.
- 9. McCord JM, Fridovich I. Superoxide dismutase: an enzymatic function for erythrocuprein. J Biol Chem 1969;244:6049.
- 10. Ohkawa H, Ohishi N, Yogi K. Assay of lipid peroxides in animal tissue by a thiobarbituric reaction. Ann Biochem 1979;95:351–8.
- 11. Oyaizu M. Studies on products of browning reaction prepared from glucose amine. Jpn J Nutr 1986;44:307–15.
- 12. Benzie IF, Strain JJ. The ferric reducing ability of plasma as a measure of "antioxidant power"-The FRAP assay. Ann Biochem 1996;239:70–6.
- 13. Dinis TCP, Madeira VMC, Almeida LM. The action of phenolic derivatives (acetaminophen, salicylate, and 5-aminosalicylate) as inhibitors of membrane lipid peroxidation and as peroxyl radical scavengers. Arch Biochem Biophys 1994;315:161–9.
- 14. Prieto P, Pineda M, Aguilar M. Spectrophotometric quantitative of antioxidant capacity through the formation of a phosphomolybdenum complex: specific application to the determination of vitamin E. Ann Biochem 1999;269:337–41.
- Naim M, Gestetner BA, Birk Y. Antioxidative and antihemolytic activities of soybean isoflavones. J Agric Food Chem 1976;24:1174–7.
- 16. Kasuga A, Aoyaga Y, Sugahara T. Antioxidant activity of fungus *Suillus bovines* (L: Fr.) O. Kuntze. J Food Sci 1995;60:1113–5.
- 17. Elmastasa M, Isildaka O, Turkekulb I, Temura N. Determination of antioxidant activity and antioxidant compound in wild edible mushrooms. J Food Compost Anal 2007;20:337–45.
- Gezer K, Duru ME, Kivrak İ, Turkoglu A, Mercan N, Turkoglu H, *et al.* Free radical scavenging capacity and antimicrobial activity of wild edible mushroom from Turkey. Afr J Biotechnol 2006;5:1924–8.
- Banerjee S, Sanjay KR, Chethan S, Malleshi NG. Finger millet (*Eleusine coracana*) polyphenols: Investigation of their antioxidant capacity and antimicrobial activity. Afr J Food Sci 2012;6:362–74.
- 20. Gutteridge MC. Reactivity of hydroxyl and hydroxyl-like radicals discriminated by the release of thiobarbituric acid reactive material from deoxy sugars, nucleosides, and benzoate. J Biochem 1984;224:761–7.
- 21. Spencer D, James EK, Ellis GJ, Shaw JE, Sprent JI. Interactions between rhizobia and potato tissue. J Exp Bot 1994;45:1475–82.
- Halliwell B, Gutteridge JMC. Free radicals in biology and medicine. 3<sup>rd</sup> Edn. Oxford: Clarendon Press; 1999. p. 936.
- Manian R, Anusuya N, Siddhuraju P, Manian S. The antioxidant activity and free radical scavenging potential of two different solvent extracts of *Camellia sinensis* (L.) O. Kuntz, *Ficus bengalensis* L. and *Ficus racemosa* L. Food Chem 2008;107:1000–7.
- 24. Menaga D, Rajakumar S, Ayyasamy PM. The free radical scavenging activity of methanolic extract of *Pleurotus florida* mushroom. Int J Pharm Pharm Sci 2013;5:601–6.
- Baskar R, Lavanya R, Mayilvizhi S, Rajasekaran P. Free radical scavenging activity of antitumour polysaccharide fractions isolated from *Ganoderma lucidum* (Fr.) P. Karst. Nat Prod Radiance 2008;7:320–5.

- Ilaentic A, Moncada S, Rosa DM. Modulation of adjuvant arthritis by endogenous nitric oxide. Br J Pharmacol 1993;110:701–6.
- 27. Sainani GS, Manika JS, Sainani RG. Oxidative stress: a key factor in the pathogenesis of chronic diseases. Nucl Med 1997;1:1.
- Lata H, Ahuja GK. The role of free radicals in health and disease. Indian J Physiol Allied Sci 2000;57:124.
- 29. Khatri DK, Manohar K, Juvekar A. Preliminary phytochemical and antioxidant evaluation of a polyherbal formulation (*Madhumi*). Int J Phytopharmacol 2013;4:322–8.
- Kokila N, Radha R, Jayshree N. *In vitro* antioxidant and antiarthritic activity of polyherbal formulation. J Pharmacogn Herb Formula 2013;13:10–5.
- Karthika C, Chitra M, Radhika K. Protective activity of Shorea robusta leaf against oxidative stress in rats. Int J Pharm Sci Res 2013;4:4754–7.
- 32. Ruch RT, Cheng SJ, Klaunig JE. Spin trapping of superoxide and hydroxyl radicals. Method Enzymol 1984;105:198–209.
- Siddhuraju P, Becker K. The antioxidant and free radical scavenging activities of processed cowpea (*Vigna unguiculata* L.) seed extracts. Food Chem 2007;101:10–9.
- Braugghler JM, Duncan CA, Chase LR. The involvement of iron in lipid peroxidation, the importance of ferrous to ferric iron ion initiation. J Biol Biochem 1986;261:10282–9.
- Khanam S, Shivprasad HN, Devi K. *In vitro* antioxidant screening models: a review. Indian J Pharm Educ Res 2004;38:180–3.
- Selvi S, Umadevi P, Devipriya D, Chinnaswamy P. In vitro antioxidant and anti lipid peroxidative potential of Calocybe indica. J Nat Rem 2010;10:27–31.
- Meir S, Kanner J, Akiri B, Hadas SP. Determination and involvement of aqueous reducing compounds in oxidative defense systems of various senescing leaves. J Agric Food Chem 1995;43:1813–9.
- Oliveira MT, Rego AC, Macedo TRA, Oliveira R. Drugs of abuse induces apoptotic features in PC12 cells. Ann N Y Acad Sci 2003;1010:667–70.
- Zou Y, Lu Y, Wei D. Antioxidant activity of a flavonoid-rich extract of *Hypericum perforatum* L. *in vitro*. J Agric Food Chem 2004;52:5032–9.
- 40. Chung KH, Hart CC, Al-Bassam S, Avery A, Taylor J, Patel PD, *et al.* Polycistronic RNA polymerase II expression vectors for RNA interference based on BIC/miR-155. Nucleic Acids Res 2006;34:1–14.
- 41. Nabavi SM, Ebrahimzadeh MA, Nabavi SF, Jafari M. Free radical scavenging activity and antioxidant capacity of *Eryngium caucasicum* Trautv and *Froripia subpinnata*. Pharmacol 2008;3:19–25.
- Kekuda TRP, Vinayaka KS, Swathi D, Suchitha Y, Venugopal TM, Mallikarjun N. Mineral composition, total phenol content and antioxidant activity of a Macrolichen *Everniastrum cirrhatum* (Fr.) Hale (Parmeliaceae). E J Chem 2011;8:1886–94.
- Yildirima NC, Turkoglub S, Yildirima S, Inceb OK. Antioxidant properties of wild edible mushroom *Pleurotus eryngii* collected from the tunnel province of Turkey. Digest J Nanomater Biostructures 2012;7:1647–54.
- Ema A, Sathish Kumar M, Jeyanthi LR, Sindhu S, Anbarasi P, Sagadevan, *et al.* Evaluation of the antiproliferative effect of *Grewia hirsuta* on HepG2 cell lines. J Acad Ind Res 2013;2:1–5.
- 45. Goodman KT, Scott PM. Risk assessment of mycotoxin ochratoxin A. Biomed Environ Sci 1989;2:179–48.
- Ebrahimzadeh MA, Pourmorad F, Bekhradnia AR. Iron chelating activity screening, phenol and flavonoid content of some medicinal plants from Iran. Afr J Biotechnol 2008;7:3188–92.
- 47. Devjani C, Verma RJ. Ameliorative effect of *Emblica officinalis* aqueous extract against ochratoxin induced lipid peroxidation in the kidney and liver of mice. Int J Occupational Med Environ Health 2010;23:1–11.

## How to cite this article

 Madhaiyan Prabu, Renganathan Kumuthakalavalli. Antioxidant activity of oyster mushroom (*Pleurotus Florida* [Mont.] Singer) and milky mushroom (*Calocybe Indica* P and C). Int J Curr Pharm Res 2016;8(3):48-51