Original Article

ANTI-DIABETIC ACTIVITY OF *LINARIA RAMOSISSIMA* (WALL) JANCH IN STREPTOZOTOCIN INDUCED DIABETIC RATS

BHAVINI I. PATEL*, PUNAM D. SACHDEVA

A R College of Pharmacy and G. H. Patel Institute of Pharmacy, Vallabh Vidhyanagar, Gujarat. Email: bhavini.patel24@gmail.com

Received: 21 June 2014 Revised and Accepted: 21 Jul 2014

ABSTRACT

Objective: To evaluate the effectiveness of Linaria ramosissima (Wall.) Janch as an antidiabetic agent in streptozotocin induced diabetic rats.

Methods: Diabetes was induced in rats by single intra-peritoneal injection of streptozotocin(STZ) 60 mg/kg body weight. Animals with BGL more than 200 mg/dl, 48 hrs after STZ injection were considered as diabetic and were used for experiment. Hydroalcoholic extract of *Linaria ramosissima* (Wall.)Janch was administered for 21 days. During treatment period daily food and water intake and weekly body weight and BGL were checked in each group of rats. Histopathology of pancreas was performed at the end of experiment.

Results: Mean food and water intake in diabetic group of rats increased as compared to normal control group. Administration of test drug (200mg/kg & 400mg/kg) significantly reduced water intake without significant reduction in food intake as compared to normal control group. Significant decrease in body weight was observed in diabetic group of rats, but administration of extract significantly improved the body weight. Elevated BGL observed in diabetic animals was significantly reduced by administration of extract.Blood glucose lowering effect of extract was confirmed by results of histopathology of pancreas.

Conclusion: The results of the studies confirmed the ethanomedicinal use of *Linaria ramosissima* (Wall.) Janch in treatment of diabetes by reducing BGL, improving parameters like food and water intake, body weight and regeneration of the islets of langerhans.

Keywords: Anti-diabetic, Glibenclamide, Blood glucose, Acute oral toxicity, Histopathology.

INTRODUCTION

Diabetes is the most important non infective epidemic to hit the globe in the present millennium [1]. The American Diabetes Association 2011 defines Diabetes Mellitus (DM) as a wide spread metabolic disease characterized by hyperglycemia and carbohydrate, protein and fat metabolism disturbances [2]. It is characterized by hyperglycemia arising as a consequence of a relative or absolute deficiency of insulin secretion, resistance to insulin action or both [3]. It is a metabolic syndrome characterized by hyperglycemia, polyuria and weight loss, inspite of polyphagia, glycosuria, ketosis and acidosis [4].

Worldwide the prevalence of diabetes mellitus is estimated to be 2.8% and is set to rise to 4.4% by 2030 [5]. According to a projection of the International Diabetes Federation (IDF), approximately 366 million people are living with diabetes and this figure is projected to increase to 552 million by the year 2030 [6], with the greatest number of cases being expected in China and India [7]. The IDF estimates the total number of diabetic subjects to be around 40.9 million in India and this is further set to rise to 69.9 million by the year 2025 [8]. India has thus been declared by WHO as the "Diabetes capital of the world" [9].

Managemnet of hyperglycemia in most patients with Type II DM should begin with lifestyle modification (diet and activity). However, diet and exercise fail to achieve glycemic control in most patients and thus pharmacologic intervention with oral hypoglycemic agents is necessary [10].

Type 2 Diabetes Mellitus patients are usually treated with oral medications including Sulfonylureas, Biguanides Alpha-glucosidase inhibitors and Thiazoliddinediones [11]. However, many of these oral hypoglycemic agents usually produce serious side effects including hypoglycemia, drug-resistance, dropsy and weight gain [12].

Hence despite the presence of various antidiabetic medicines on the market, diabetes and related complications continue to be a major problem. Traditionally used medicinal plants can provide an alternative approach to treat diabetes [13], because of their effectiveness, less side effects and relatively low cost [14].

The ethanobotanical reports state that about 800 plants may possess antidiabetic potential [15]. WHO has recommended that traditional plant treatments for diabetes warrant further evaluation [16], hence over 150 plant extracts and some of their active principles, including flavanoids, tannins, alkaloids etc., are used for treatment of diabetes [17]. Although several medicinal plants have gained importance for the treatment of diabetes mellitus, many remain to be scientifically investigated [18].

The plant Linaria ramosissima (Wall.)Janch (syn: Kickxia ramosissima). belongs to family Scrophulariaceae. It is commonly called "kanodi" and "Bhintgalodi" in Gujarati. It is distributed throughout India, usually in rocky and stony places, Ceylon and Upper Burma [19]. It is also found in the saurashtra region of Gujarat [20]. Medicinal properties like Mutrala (diuretic), Rechak (purgative), Tikta (bitter), Raktapittahara (blood disorders) have been reported [20]. Folk people in saurashtra region use this plant to treat urinary stone [20]. Leaves mixed with black pepper are given in fever, Root is used in the management of snake and scorpion bite [21]. Traditionally the plant has been reported as an effective remedy for diabetes. However literature survey shows no scientific evidence regarding antidiabetic activity of the plant.

Hence the present study has been undertaken with the aim to evaluate the antidiabetic activity of plant of Linaria ramosissima (Wall.) Janch.

MATERIALS AND METHODS

Plant material and Preparation of extract:

Whole plant of *Linaria ramosissima* (Wall.)Janch was collected from Gujarat Ayurvedic University, Jamnagar and was identified and authenticated by Dr. D.B.Patel, Professor & Head, Department of Botany, Bansilal Amrutlal College of Agriculture (B.A.C.A), Anand. **(Authentification no: BACA/GPB/623/13)** A herbarium of whole plant was prepared and deposited in Pharmacognosy Deparatment

of A.R.College of Pharmacy, Vallabh Vidyanagar (Herbarium no: BIP/LR-25/2/ARGH-14)

Plant material was dried in shade at room temperature for 15 days, coarsely powdered and the powder was passed through 40# sieve. The powdered plant material was subjected to soxhlet extraction using water:methanol (30:70) to obtain hydroalcoholic extract. Hydroalcoholic extract was evaporated under reduced pressure at low temperature (30°C) to dryness to yield a brownish green colour residue. (yield- 13.64% w/w) The residue obtained was stored in an airtight glass container and was used for evaluation of antidiabetic activity.

Phytochemical analysis

The dried extract was subjected to qualitative analysis for the detection of various phytoconstituents present.

Animals

Albino rats of wistar strain, weighing 200-300g were procured from Zydus Research Centre, Ahmedabad. The animals were housed under standard conditions i.e $26 \pm 2^{\circ}$ C temp. and relative humidity 44-56%, maintained on a 12 hr light/dark cycle and had free access to food and water. The animals were acclimatized to the laboratory environment 1 hr before the experiments. All experiments were conducted during the light period (08.00-16.00hrs.). The protocol (No: CPCSEA/IAEC/AR CP/13-14/01) of the study was approved by the Institutional Animal Ethical Committee (IAEC) and experiments were conducted according to the guidelines of CPCSEA (Committee for the Purpose of Control and Supervision of Experiments on Animals).

Acute oral toxicity study [22]

Acute oral toxicity study was performed as per Organization for Economic Cooperation and Development (OECD) guidelines 423. Stepwise dose of extract of *Linaria ramosissima* (Wall.)Janch (5,50,300 and 2000 mg/kg body weight p.o.),was administered and animals were observed individually atleast once during the first 30 minutes and periodically during the first 24 hrs., with special attention given during the first 4 hrs and daily thereafter, for a total of 14 days.

Drugs

Streptozotocin was purchased from Himedia Chemical, Mumbai, India. Glibenclamide was obtained as gift sample from Yarrow Chem Products, Mumbai, India.

Induction of Diabetes [23]

Diabetes was induced in the overnight fasted rats by a single intraperitoneal injection of 60 mg/kg body weight streptozotocin dissolved in 0.1 M sodium citrate buffer, pH 4.5. After the injection they had free access to food and water. The animals were allowed to drink 5% glucose solution overnight to overcome hypoglycaemic shock. The development of diabetes was confirmed after 48 hrs of streptozotocin injection. The animals having blood glucose level more than 200 mg/dl were considered as diabetic and were used for the experimentation.

Experimental procedure

In experiment total 30 rats were used. The rats were divided into 5 groups of 6 rats each

GROUP I: Normal control rats

GROUP II: Diabetic control rats

GROUP III: Diabetic rats treated with standard drug (Glibenclamide 10 mg/kg b.wt, po)

 $\ensuremath{\mathsf{GROUP\,IV}}$: Diabetic rats treated with plant extract (200mg/kg b.wt, po)

 $\ensuremath{\mathsf{GROUP}}\xspace V$: Diabetic rats treated with plant extract (400mg/kg b.wt, po)

All drug treatment was given for 21 days. During treatment period daily food & water intake of rats in each group was checked. Body

weight of rats of all groups was checked at weekly interval. Blood sample was collected from retro orbital plexus on day 0 and on 7th, 14th and 21st day after drug treatment to estimate blood glucose level. On the last day of experiment (21st day of drug treatment) 3 rats from each treatment group were sacrificed & specimens of pancreas were obtained for histopathological studies.

Statistical Analysis

All the results were tested for significance using One-Way ANOVA followed by Tukey's test and the results were expressed as mean \pm S.E.M. at the probability level of 95%.

RESULTS

Phytochemical analysis

Preliminary qualitative phytochemical screening showed the presence of Carbohydrates, Phytosterol, Cardiac glycosides, Saponins, Flavonoids, Alkaloids, Tannins and Phenolic compounds.

Acute oral toxicity study

The acute oral toxicity study showed that hydroalcoholic extract of *Linaria ramosissima* (Wall.)Janch was devoid of any toxicity even at the dose of 2000 mg/kg b.wt. Hence 200mg/kg and 400 mg/kg b.wt doses were selected for the study.

Effect of Hydroalcoholic extract of *Linaria ramosissima* (Wall.)Janch on food intake

Mean food intake in diabetic group of rats increased from 78.18±8.91 gms in the 1st week to 101.7±3.92 gms (+30%) at the end of 3rd week. Treatment with standard antidiabetic drug Glibenclamide (10mg/kg) reduced mean food intake from 101.7±16.8 gms in the 1st week to 79.87±17.32 gms (-21.4%) by the end of 3rd week. Similarly both doses of test drug (200mg/kg & 400 mg/kg) reduced mean food intake from 96.58±6.57 gms amd 91.45±5.23 gms in the 1st week to 83.28±16.15 gms (-13.75%) and 64.11±4.59 (-29.89%) gms at the end of 3rd week respectively. However as compared to normal control group the decrease in food intake produced by standard and test drugs was not significant.(Fig 1)

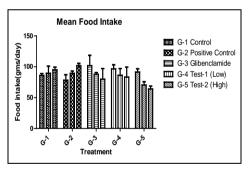


Fig. 1: Mean food intake per week up to 3 weeks in different treatment groups

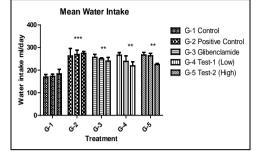
Effect of Hydroalcoholic extract of *Linaria ramosissima* (Wall.)Janch on water intake

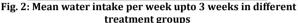
Water consumed by the different groups of rats was recorded daily and mean water intake per week was calculated. Water intake in diabetic group of rats increased from 262.6 ± 33.11 ml in the 1st week to 273.7 ± 8.41 ml (+4.2%) in the 3rd week. Comparing the water intake in diabetic groups with normal control animals the difference was found to be highly significant(p<0.001). **(Fig-2)**

The mean water consumption in Glibenclamide treated group of rats was 256.1 ± 14.08 ml in the 1st week which reduced to 239.9 ± 16.4 ml (-6.32%) in the 3rd week. Similarly with both doses of test drug (200mg/kg & 400 mg/kg) the mean water consumption reduced from 266.3 ± 11.22 ml and 266.9 ± 11.52 ml in the 1st week to 218.9 ± 18.02 ml (-17.7%) and 24.7 ± 4.84 ml (-15.8%) in the 3rd week respectively. Comparing the decrease in mean water intake

produced by standard drug Glibenclamide and both doses of test drug with normal control (non-diabetic)animals the difference was found to be significant(p<0.01).

However, comparing the decrease in water consumption produced by both standard and test drugs with positive control (diabteic) groups of animals the difference was not found to be significant.





Effect of Hydroalcoholic extract of *Linaria ramosissima* (Wall.)Janch on Body Weight

Table 1 shows the effect of hydroalcoholic extract of *Linaria* ramosissima (Wall.)Janch on body weight of the animals.

As is evident from the table, mean body weight of diabetic control animals on 0 day was 220±9.3 gms which reduced to 178±6.0 gms (-19.%) on 21st day of induction of diabetes. This decrease in body weight of diabetic animals was statistically significantly (p<0.001) as compared to normal control animals. Administration of hydroalcoholic extract of *Linaria ramosissima*(Wall.)Janch in both doses (200mg/kg & 400mg/kg) showed significant (p<0.05) improvement in body weight compared to diabetic groups. Mean body weight of animals in test group-1 and test group-2 increased from 216.7±12.8 gms & 220±9.3 gms on 0 day to 240.8±12.19 gms (+11.1%) and 245±7.63 gms (+11.3%) on 21st day after induction of diabetes respectively.

Similar results of significant improvement (p<0.01) in body weight as compared to diabetic control group of animals were obtained with administration of standard antidiabetic drug Glibenclamide (10 mg/kg), which increased body weight from 228.3 ± 10.14 gms to 251.7 ± 13.08 gms (+10.2%) at the end of the study (21st day).

Groups	Mean Body weight (gms)				
	0 day	7 day	14 day	21 day	
Normal control	236.7±7.6	255.0±8.0(+7.7%)	259.2±7.4(+9.5%)	265.0±9.8(11.9%)	
Positive control	220.0±9.3	203.3±8.0 (-7.5%)	190.8±7.5 (-13.2%)	178.0±6.0(-19.0%)***	
Standard	228.3±10.1	240.0±29.6 (+5.1%)	242.5±12.3 (+6.21%)	251.7±13.08 (+10.2%)**	
(Glibenclamide 10mg/kg)					
Test extract	216.7±12.8	221.7±14.3(+2.3%)	230.0±12.3 (6.1%)	240.8±12.1 (+11.1%)*	
(200mg/kg)					
Test extract	220.0±9.3	225.8±8.4(+2.6%)	229.2±8.7(+4.1%)	245.0±7.63(+11.3)*	
(400mg/kg)					

Values are expressed as mean ± S.E.M., n=6, ***p<0.001,as compared to normal control group **p<0.01, *p<0.05, as compared to positive control group by One Way ANOVA followed Tukey's test

Time dependent effect of *Linaria ramosissima* (Wall.)Janch on blood glucose level (BGL)

Table 2 shows variation in BGL from 1st day to 21st day in each group. Mean BGL in non-diabetic control animals ranged from 94.33 \pm 1.68 mg/dl on 0 day to 98.50 \pm 0.99 mg/dl on 21st day of the study. In STZ-induced diabetic animals the BGL increased significantly (p<0.001) from 447.2 \pm 31.32 mg/dl on 0 day to 530.2 \pm 24.22 mg/dl (+18.5%) on day 21st day of the study. With

standard drug Glibenclamide (10mg/kg) the BGL reduced from 381.2 ± 52.79 mg/dl on 0 day to 237.8 ± 37.38 mg/dl on 21^{st} day. This difference was found to be highly significant(p<0.001) when compared with diabetic control group. Test drug in the dose of 200 mg/kg significantly reduced BGL from 402.8 ± 43.4 mg/dl on 0 day to 292.7 ±39.15 mg/dl (-27.3%) on 21^{st} day. 400mg/kg dose of test drug also reduced BGL from 470.0 ± 12.65 on 0 day to 305.7 ± 16.56 mg/dl (-34.9%) on 21^{st} day, but the difference was not found to be significant.

Day wise Blood Glucose Level (mg/dl)				
0 day	7 days	14 days	21 days	
94.33±1.68	95.33±1.49	97.83±0.87	98.50±0.99	
447.2±31.3	494.2±27.2(+10.5%)	512.3±26.2(+14.5%)	530.2±24.2(+18.5%)***	
381.2±52.7	342.3±51.8(-10.2%)	297.5±46.2(-21.9%)	237.8±37.3(-37.6%)***	
402.8±43.4	366.3±48.7(-9.06%)	325.5±45.4(-19.1%)	292.7±39.15(-27.3%)**	
470.0±12.6	423.7±17.8 (-9.8%)	374.8±16.6(-20.2%)	305.7±16.5(-34.9%)	
	0 day 94.33±1.68 447.2±31.3 381.2±52.7 402.8±43.4	0 day 7 days 94.33±1.68 95.33±1.49 447.2±31.3 494.2±27.2(+10.5%) 381.2±52.7 342.3±51.8(-10.2%) 402.8±43.4 366.3±48.7(-9.06%)	0 day 7 days 14 days 94.33±1.68 95.33±1.49 97.83±0.87 447.2±31.3 494.2±27.2(+10.5%) 512.3±26.2(+14.5%) 381.2±52.7 342.3±51.8(-10.2%) 297.5±46.2(-21.9%) 402.8±43.4 366.3±48.7(-9.06%) 325.5±45.4(-19.1%)	

Values are expressed as mean ± S.E.M., n=6, ***p<0.001, as compared to normal control group***p<0.001, **p<0.05, as compared to positive control group by One Way ANOVA followed by Tukey's Test

Effect of Hydroalcoholic extract of *Linaria ramosissima* (Wall.)Janch on pancreas by Histopathology

Fig 3A shows normal islets of langerhans and β cells in pancreas of normal control group.(non-diabetic) In positive control group

(Diabetic) the number of pancreatic islets as well as β -cells is reduced as compared to control group with most of β -cells being destroyed. (Fig 3B) Section of pancreas from groups treated with glibenclamide (Fig 3C) and both doses of test drug (Fig 3D & 3E)

showed increase in pancreatic islets & number of β -cells in the pancreas. The damaged β -cell seen after induction of diabetes were no longer observed after treatment with extract. This indicates that the test drug causes regeneration of β -cell of islets of langerhans of pancreas and restores normal cellular appearance and size of islets with hyperplasia.

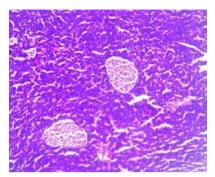


Fig. 3a: Normal Control (Non-diabetic)

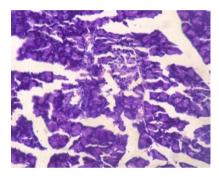


Fig. 3b: Positive Control (Diabetic)

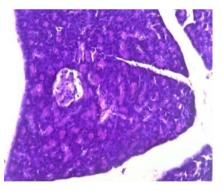


Fig. 3c: Glibenclamide Standred drug (10 mg/kg)

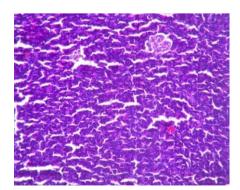


Fig. 3d: Test-1 (Low dose) 200 mg/kg

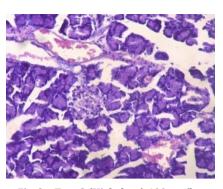


Fig. 3e: Test-2 (High dose) 400 mg/kg

DISCUSSION

Streptozotocin (STZ) is an antibiotic obtained from *Streptomyces* achromogenes. It enters the pancreatic β -cells via a glucose transporter-GLUT2 and causes alkylation of deoxyribonucleic acid (DNA) **[24]**, which causes the generation of superoxide, hydrogen peroxide, nitric oxide and hydroxyl radicals which are responsible for β -cell damage and necrosis resulting in diabetes **[25]**.

Usually, the intraperitoneal injection of a single dose of STZ 60 mg/kg body weight exerts direct toxicity on β -cells resulting in necrosis within 48 hrs and causes permanent hyperglycemia **[26]**. STZ was used in the present study for the induction of diabetes in rats.

Diabetes is characterized by polyphagia which is evident from the increase in food intake observed in diabetic group of animals. However, the food intake in animals treated with extract was less as compared to diabetic control group and was comparable with standard antidiabetic drug Glibenclamide.

Another characteristic sign of diabetes is polydypsia which is evident by increase in water intake in diabetic groups of animals. The plant extract decreased the water consumption to near normal levels, which were comparable to standard antidiabetic drug Glibenclamide. Other scientists have also reported decrease in food and water intake by use of herbal antidiabetic drugs [27].

Deficiency of insulin brings about improper metabolism of carbohydrate, lipids and proteins leading to increased gluconeogenesis, glycogenolysis, lipolysis and muscle wasting **[28]**. The phenomenon of muscle wasting occurs due to nonavailability of carbohydrate for energy metabolism leading to protein breakdown in skeletal muscles and thus causes loss of body weight in diabetic animals **[29]**.

Treatment with the extract showed a significant increase in body weight, which may be due to the effectiveness of the drug in reversing gluconeogenesis and improving glycemic control and thus exert a protective effect in controlling muscle wasting **[27]**.

Similar observations of increase in body weight produced by various plant drugs with esthblished antidiabetic activity like *Juniperus communis*(L.) **[29]**, *Calotropis procera* **[30]**, *Garuga pinnata* **[31]**, *Cinnamomum tamala* **[32]** and *Momordica tuberose* **[33]** have been reported.

Diabetic animals showed increase in BGL which was reduced by administration of both doses of extract. The possible mechanism by which the extract produced antidiabetic glycemic action in diabetic rats may be by potentiating the insulin effect of plasma by increasing either the pancreatic secretion of insulin from the existing β -cells or by its release from the bound form **[34]** or by promoting regeneration of β -cells or inhibiting endogenous glucose production or by inhibition of intestinal glucose absorption **[35]**. A pancreatic mechanism is possible because in mild diabetes induced by STZ all the β -cells of the pancreas are not destroyed. The surviving β cells retain the capacity to synthesize and secrete insulin **[36]**.

It is reported that flavanoids [37], phytosterols [1], alkaloids [38], glycosides [39] and phenolic compounds [40, 41], constitute the

active biological principle of most medicinal plants with hypoglycemic and antidiabetic properties **[42]**. Flavonoids are reported to regenerate the damaged pancreatic β cells in diabetic animals **[43]**. Polyphenol such as tannins and saponins reduce blood glucose level through inhibition of α amylase and sucrase from the intestine **[44, 45]**. Thus the significant antidiabetic activity of the extract may be attributed to synergistic effect of such phytoconstituents present in the plant. Results of histopathological studies show increase in the number of islets and β cells through proliferation of residual, pancreatic β cells thus restoring normal structural integrity of islets of langerhans **[27]**. Various histopathology studies have demonstrated an effective increase in number of β -cells in pancreas of diabetic rats treated with various plant extracts like *Momordica tuberose* **[34]**, *Juglans regia* L **[46]** and *Teucrium polium* **[47]** all of which possess antidiabetic activity.

CONCLUSION

In conclusion, the findings of present study demonstrated that hydroalcoholic extract of *Linaria ramosissima* (Wall.)Janch possesses favourable antidiabetic activity against STZ induced diabetes in wistar rats thus confirming the traditional/ethanomedical use of the plant in the treatment of diabetes.

REFERENCES

- 1. Rao AS, Vijay Y, Deepthi T, Sri Lakshmi Ch, Rani V, Rani S *et al.* Anti diabetic effect of ethanolic extract of leaves of *Ocimum sanctum* in alloxan induced diabetes in rats. Int J Basic and Clin Pharmacol 2013;2:613-6.
- Wan LS, Chen CP, Xiao ZQ, Wang YL, Min QX, Yue Y et al. In vitro and in vivo anti-diabetic activity of Swertia kouitchensis extract. J Ethnopharmacol 2013;3:622-30.
- 3. Weng Y, Yu L, Cui J, Zhu YR, Guo C, Wei G *et al.* Antihyperglycemic, hypolipidemic and antioxidant activities of total saponins extracted from *Aralia taibaiensis* in experimental type 2 diabetic rats. J Ethnopharmacol 2014;2:553-60.
- Jain A, Jasmine, Sharma S, Saini V. Hypolipidemic, Hypoglycemic and Antioxidant potential of Saraca asoca ethanolic leaves extract in streptozotocin induced experimental diabetes. Int J Pharma and Pharma Sci 2013;5:302-05.
- Kavitha KN, Dattatri AN. Experimental Evaluation of antidiabetic activity of *Swertia chirata*-Aqurous Extract. J Pub Health Med Res 2013;1:71-5.
- 6. Ibrahim MA, Islam MS. Anti-diabetic effect of the acetone fraction of *Senna singueana* stem bark in a type 2 diabetes rat model. J Ethnopharmacol 2014;2:326-35.
- Asha B, Krishnamurthy KH, Siddapadevaru. Evaluation of antihyperglycemic activity of *Zingiber officinale* in albino rats. J Chem Pharma Res 2011;3:452-56.
- Sicree R, Shaw J, Zimmet P. Diabetes and impaired glucose tolerance. 3 rd ed. Belgium:International Diabetes Federation;2006.
- Gowri K, Elanchezhiyan C, Hemalatha S, Sartaj AA. Effect of Naringi crenulata extract on histopathological changes in streptozotocin induced diabetic rats. Asian J Sci Technology 2013;4:136-39.
- 10. Nathan DM, Buse JB, Davidson MB, Ferrannini E, Holman RR, Sherwin R. Medical management of hyperglycemia in type 2 diabetes a consensus algorithm for the initiation and adjustment of therapy:a consensus statement of the American Diabetes Association and the European Association for the Study. J Diabetes. Diabetes Care 2009;32:193-203.
- 11. http://www.en.wikipedia.org/wiki/Diabetes_mellitus
- Tahrani AA, Piya MK, Kennedy A, Barnett AH. Glycaemic control in type 2 diabetes targets and new therapies. J Pharmacol Ther 2010;125:328-61.
- 13. 13.Ezzat SM, Fathalla MH,Salah AG. Antihyperglycemic and antihyperlipidemic effects of the methanol extracts of *Cleome ramosissima* Parl.,*Barleria bispinosa* Vahl. and *Tribulus macropterus*. J Boiss Bulletin of Faculty of Pharmacy Cairo University 2013:1-7.
- Venkatesh S, Reddy GD, Reddy BM, Ramesh M, Rao AV. Antihyperglycemic activity of *Caralluma attenuata*. J Fitoterapia 2003;74:274-9.

- Yasodamma N, Shanthi KS Alekhya C. Antidiabetic activity of Sebastiania chamaelea Muell.ARG. leaf extracts in alloxan induced diabetic albino rats. Int J Pharm Pharm Sci 2013;5:577-83.
- Senthilkumar MK, Sivakumar P, Faisal Changanakkattil, Rajesh V, Perumal P. Evaluation of Anti-diabetic activity of *Bambusa vulgaris* leaves in Streptozotocin induced Diabetic rats. Int J Pharm Sci Drug Res 2011;3:208-10.
- Adaramoye OA, Adeyemi EO. Hypoglycaemic and Hypolipidaemic effects of fraction from kolsviron, a biflavonoid complex from *Garcinia Kola* in streptozotocin-induced diabetes mellitus rats. J Pharm Pharmacol 2006;58:121-26.
- Punitha R, Vasudevan K, Manoharan S. Effect of *Pongamia pinnata* flowers on blood glucose and oxidative stress in alloxan induced diabetic rats. Indian J Pharmacol 2006;38:62-63.
- 19. Kirtikar KR, Basu BD. Indian medicinal Plants. 2nd ed. Dehra Dun:International book distributors;2005.
- 20. Pandya PN, Aghera HB, Ashok BK. Pharmacological study of Diuretic activity of *Linaria ramosissima* (Wall.) Janch leaves in albino rats. J AYU 2012;33:576-8.
- 21. Bole P V, Pathak JM. Flora of Saurashtra. The Director Botanical Survey of India, New Delhi:Deep Printer;1988.
- OECD, Guideline 423 Acute oral toxicity:Environmental Health and Safety Monograph series on Testing and Assessment No.24 (2000)
- 23. Rajalakshmi M. Anti-diabetic properties of *Tinospora cordifolia* stem extracts on streptozotocin induced diabetic rats. Afr J Pharm Pharmacol 2009;5:171-80.
- 24. Das PK, Bhattacharya S, Pandey JN, Biswas M. Antidiabetic activity of *Trapa natans* fruit peel extract against streptozotocin induced diabetic rats. Global J Pharmacol 2011;5:186-90.
- 25. Szkudelski T. The mechanism of alloxan and streptozotocin action in β -cell of the rat pancreas. J Physiol Res 2001;50:536-46.
- Daisy P, Jeevakani FG. Evaluation of Antidiabetic activity of various extracts of *Cassia auriculata* Linn. bark on streptozotocin induced diabetic wistar rats. Int J Pharm Pharm Sci 2012;4:312-8.
- Ahmed F, Urooj A. Antidiabetic activity of *Ficus glomerata* stem bark in streptozotocin-induced diabetic rats. Global J Pharmacol 2008;2:41-5.
- Cohen RM, Haggerty S, Herman WH. HbA1c for the diagnosis of diabetes and prediabetes is it time for a mild-course correction. J Clin Endocrinol Metab 2010;95:5203-06.
- Banerjee S, Singh H, Chatterjee TK. Evaluation of antidiabetic and antihyperlipidemic potential of methanolic extract of *Juniperus communis*(L.) in streptozotocin-nicotinamide induced diabetic rats. Int J Pharm Bio Sci 2013;4:10-7.
- Roy S, Sehgal, Padhy BM, Kumar VL. Antioxidant and protective effect of latex of *Calotropis procera* against alloxan-induced diabetes in rats. J Ethnopharmacol 2005;102:470-73.
- Shirwaikar A, Rajendran K, Barik R. Effect of aqueous bark extract of *Garuga pinnata* Roxb. in steptozotocin-nicotinamide induced type-II diabetes mellitus. J Ethnopharmacol 2006;107:285-90.
- Chakraborty U, Das H. Antidiabetic and antioxidant activities of *Cinnamomum tamala* leaf extracts in STZ-treated diabetic rats. Global J Biotech Biochem 2010;5:12-18
- Palanivel V, Mohammed S, Senthil kumar KL. Antidiabetic and hypolipidemic activities of *Momordica tuberosa* unripe fruit extract on diabetic induced rats. Int J Advanced Pharm Genuine Res 2013;1:33-40.
- 34. Rahman NN, Khan M, Hasan R. Bioactive components from *Ficus glomerata*. J Pure and Applied Chemistry 1994;66:2287.
- 35. Venkateshwarlu E, Sharvana Bhava BS, Reddy PR, Mahathi K. Evaluation of antidiabetic and hypolipidemic activity of *Pseudarthria viscida* (whole plant) in streptozotocinnicotinamide induced type II diabetic rats. Global J Pharmacol 2013;7:192-7.
- 36. Onoagbe IO, Negbenebor EO, Ogbeide VO, Dawha IH, Attah V, Omonkhua AA. A study of the antidiabetic effects of *Urena lobata* and *Sphenostylis stenocarpa* in Streptozotocin induced diabetic rats. European J Scientific Res 2010;43:6-14.
- Gupta R, Kumar D, Chaudhary AK, Singh R, Maithani M. Antidiabetic activity of *Passiflora incarnata* Linn. in streptozotocin induced diabetes in mice. J Ethnopharmacol 2012;139:801-06.

- Loew D and Kaszkin M. Approaching the problem of bioequivalence of herbal medicinal products. Phytother Res 2002;16:705-11.
- Jaiswal D, Rai PK and Watal G. Antidiabetic effect of Withania coagulans in experimental rats. Indian J Clin Biochem 2009;24:88-93.
- Wollenweber LE, Cody V, Middletone EJ, Harborne JB, Bertz A. Plant flavonoid in biological and medicinal:biochemical, cellular and medicinal properties. Progress in Clin Biological Res 1988;280:1-461.
- Saraf S, Ashawat MS, Saraf S. Flavonoids:a nutritional protection against oxidative and UV induced cellular damages. Phcog Rev 2007;1:30-40.
- 42. Bhattacharya S. Are we in the polyphenol era ? J Phcog Res 2011;3:47.

- 43. Siyem D, Syngai G, Khup PZ, Khongwir BS, Kayang H. Hypoglycemic effects of *Potentilla fulgens* Lin normal and alloxan diabetic mice. [Ethnopharmacol 2002;83:55-61.
- 44. Chakravarthy BK, Gupta S, Gode KH. Functional cell regeneration in the islets of pancreas in alloxan induced diabetic rats by (-) epicetechin. J Life Science 1982;13:2693-97.
- Emilien G, Maloteaux JM, Ponchon M. Pharmacological management of diabetes:recent progress and future perspective in daily drug treatment. J Pharmacol Ther 1999;81:37-51.
- 46. Jelodar G, Mohsen M, Shahram S. Effect of *juglans regia* (walnut leaf)L.,Coriander and Pomegranate on blood glucose and histopathology of pancreas of alloxan induced diabetic rats. Afr J Trad CAM 2007;4:299-305.
- 47. Yazdanparast R, Eslami MA, Ashrafi HJ. *Teucrium polium* extract effects pancreatic function of streptozotocin diabetic rats. Iranian Biomed J 2005;9:81-5.